

Open Access

Research Article

The Anti-Inflammatory Meroterpenes

Sibel Avunduk

Vocational School of Health Care, Mugla University, Marmaris, Mugla, 48187 Turkey.

Article Info

Received: March 07, 2022 Accepted: March 16, 2022 Published: March 22, 2022

*Corresponding author: Sibel Avunduk, Vocational School of Health Care, Mugla University, Marmaris, Mugla, 48187 Turkey.

Citation: Sibel Avunduk (2022) "The Anti-Inflammatory Meroterpenes". J Pharmacy and Drug Innovations, 3(4); DOI: http://doi.org/03.2022/1.1048.

Copyright: © 2022 Sibel Avunduk. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract

The meroterpenes are one class of the natural product bearing the terpenoid structure. They have an excellent potential for various biological activities. Inflammation is a serious problem that many people face as a complication of any chronic disease or disease itself. Therefore, we have scanned the literature to compile the research about whether the meroterpenes have potential candidates for the anti-inflammatory drug. It has been mentioned many meroterpenes have IC50 values in the μ M band. So, they are very active compounds according to anti-inflammatory assays tested.

Inflammation is a main immune reaction that serves survival during infection or injury and provides tissue homeostasis under a kind of risky conditions. Inflammation comes at the loss of a temporary reduction in tissue role, which can consecutively assist in the pathogenesis of diseases of changed homeostasis [1].

Inflammation is observed as the main complication in bacterial, viral, fungal, or parasitic infections; in anaphylaxis; in environmental diseases emerged with smoke inhalation, asbestos exposure, etc. like reasons, in autoimmune diseases such as rheumatoid arthritis, gout, and intestinal diseases; as well as in chronic diseases such as diabetes [2].

On the other hand in the last thirty years, it has also become clear that a much larger type of disease has revealed cellular and molecular proof for inflammation. These contain chronic arterial and venous disease [3,4], myocardial ischemia [5–7], acute cerebral stroke and Alzheimer's chronic disease (8-13), and more currently arterial hypertension [14] and cancer [15–17].

Nuclear factor-kB (NF-kB) is an essential regulator of inflammation, and activation of NF-kB stimulates inflammation-related metabolic disorders such as obesity, type 2 diabetes, and atherosclerosis [18]. Macrophages, as elemental immune cells, have been identified as essential effector cells in the starting and development of inflammation and insulin resistance [19]. Lots of papers exist using Nuclear factor-kB (NF-kB) assay as anti-inflamatory activity. Hyperinoids A and B, two polycyclic meroterpenoids from Hypericum patulum were investigated for the inhibitory activities in NF-kB pathway luciferase assay and the effects on the LPS-induced inflammatory responses in macrophages by Jia and the research group [20]. They have found Hyperinoids A and B showed powerful inhibitory activities in NF-kB pathway luciferase assay with IC50 values of 0.75±0.17 and 1.19 ±0.48 mmol/L, respectively. In this study, bortezomib (PS- 341) was utilized as the positive control (IC50 = 0.07 ± 0.01 mmol/L). Additionally, they were checked for influences on the LPS-activated inflammatory responses in RAW 246.7 macrophages and main mouse BMDM cells. The MTS assay demonstrated no noticeable cytotoxicity against RAW 246.7 cells at the analyzed concentrations. The mRNA levels of some pro-inflammatory genes, like IL-1β, IL-6, and iNOS, were downregulated by Hyperinoids A and B [20].

(+)- and (-)-gancochlearols A and B, two pairs of dimeric meroterpenoid enantiomers have been isolated from the fruiting bodies of Ganoderma cochlear by Quin and his

sresearch group. Their structures have been elucidated by activity (NO-Nitric oxide) in RAW 264.7 cells (IC50 = spectroscopic techniques. Biological assays showed that the 37.69/33.76 µM) [27,29]. enantiomers of 1 and 2 are cytotoxic against three human cancer cell lines (A549, K562, Huh-7) and could inhibit COX-2 The research of a marine algae-associated fungus Pleosporales sp. (cyclooxygenase-2) expression with IC50 values less than 10 µM resulted in the obtaining of four new merosesquiterpenoids [21].

Shi et all. have isolated eight pairs of meroterpenoid enantiomers and four achiral meroterpenoids from Rhododendron anthopogonoides Maxim. Compounds (+)-anthoponoid E /(-)anthoponoid E -(+) Anthoponoid G /(-) Anthoponoid G and Anthoponoid H have been identified as NF-KB pathway inhibitors, and (+)-anthoponoid E, (-) Anthoponoid G, and Anthoponoid H further showed suppressive effects on the LPS (lipopolysaccharide)-induced inflammatory responses in RAW 264.7 macrophages [22].

Furanaspermeroterpenes A and B, with an unprecedented 6/6/6/5/5 pentacyclic skeleton, and five new analogs aspermeroterpenes D-H were co-isolated from the marinederived fungus Aspergillus terreus GZU-31-1 by Tang et all. All of the isolates were scanned on the inhibitory activities against in vitro by suppressing nitric oxide (NO) generation in lipopolysaccharide-induced nitric oxide production in RAW lipopolysaccharide-induced RAW264.7 cells with an IC50 value 264.7 cells, and aspermeroterpenes D-H displayed notable antiinflammatory activity with IC50 values ranging from 6.74 to 29.59 µM than the positive control (Indomethacin, IC50 30.98 A phytochemical investigation of the alga Cystoseira usneoides μM) [23].

meroterpenoids isolated from the brown seaweed Cystoseira usneoides have been evaluated by Zbakh and his research group. The algal meroterpenoids usneoidone Z, 11-hydroxy-1'-Omethylamentadione, cystomexicone B, cystomexicone A, 6-cisamentadione-1'-methyl ether, cystodione A, and cystodione B.were assayed for their inhibitory effects on the production of the pro-inflammatory cytokines tumor necrosis factor (TNF- α), interleukin-6 (IL-6), and interleukin-1 β (IL-1 β), and the meroterpenoids, expression of cyclooxygenase-2 (COX-2), and inducible nitric oxide synthase (iNOS) in LPS-stimulated THP-1 human macrophages. All compounds remarkably reduced the production pyran-4-yl)methyl)butoxy)-6-oxohexyl of TNF-a, IL-6, and IL-1β, and suppressed the COX-2 and iNOS (C29) were isolated from the methanol:ethyl acetate fraction of expression, in LPS-stimulated cells (p < 0.05) [24].

An uncommon austinoid, 1,2-dehydro-terredehydroaustin, has pyran-4-yl)methyl)butoxy)-6-oxohexyl been isolated from the mangrove endophytic fungus Aspergillus terreus, and its' anti-inflammatory activity was tested and the new compound displayed feable inhibition effect against the production of nitric oxide (NO) in lipopolysaccharide (LPS)induced RAW 246.7 mouse macrophages with an IC50 value of 42.3 µM [25].

In another study, six farnesyl meroterpenes, consisting of two new meroterpenoids, chrysogenester and 5-farnesyl-2-methyl-1-Omethyl hydroquinone, were found in the jellyfish-related fungus Penicillium chrysogenum J08NF-4. Chrysogenester managed to suppress the in vitro inflammatory answer by joining in the peroxidase proliferator-activated receptor (PPAR-y)/NF-kB signaling line [26,29].

Brasilianoids B-C which isolated from sponge-derived fungus Penicillium brasilianum exhibit moderate anti-inflammatory Aspertetranones A-D was obtained from the marine algal-related

(pleosporallins A-D); pleosporallins A-C showed average antiinflammatory against the lipopolysaccharide (LPS)-induced intracellular IL-6 production of murine macrophage cell line RAW264.7 [28, 29].

Berkeleyacetal C (BAC) isolated from Penicillium sp. from a soil sample collected in Fukushima, suppressed NO production and induction of iNOS protein in RAW264.7 cells triggered by the Toll-like receptor (TLR) 2 ligand, peptidoglycan (PGN) or TLR4 ligand, lipopolysaccharide (LPS). BAC inhibits LPS- and PGNinduced NO generation and iNOS expression by lessening the level of the translocating of NF-kB in nuclear through preventing the kinase activity of IRAK-4 in inflammatory cells. [30]

Amestolkolides A-D was isolated from the mangrove endophytic fungus Talaromyces amestolkiae YX1 by Chen's research group. Amestolkolide B displayed powerful anti-inflammatory activity of 1.6 ± 0.1 mM [31].

has come out to obtaining the of six new meroterpenoids, cystodiones A-F, and also six known derived compounds: 6-cis-The anti-inflammatory and anticancer properties of eight amentadione-1'-methyl ether, amenta- dione-1'-methyl ether, cystomexicone A, cystomex- icone B, usneoidone Z, and its corresponding 6E isomer. In anti-inflammatory tests, usneoidone Z and its' 6E isomer exhibited remarkable activity as inhibitors of the generation of the proinflammatory cytokine TNF- α in LPSinduced THP-1 human macrophages [32].

> Three and anti-inflammatory oxygenated antioxidant 1-(3-methoxypropyl)-2-propylcyclohexane (C13), 3-(methoxymethyl)heptyl 3-(cyclohex-3-enyl) propanoate (C18) , and 2-ethyl-6-(4-methoxy-2-((2-oxotetrahydro-2H-5-ethyloct-4-enoate red seaweed Kappaphycus alvarezii by Makkar & Chakraborty. The compound 2-ethyl-6-(4-methoxy-2-((2-oxotetrahydro-2H-5-ethyloct-4-enoate showed strong in vitro inhibitory activities against proinflammatory 5- lipoxidase (IC50 1.04 mg/mL), which pointed out its potential anti-inflammatory features against promoted inflammatory mediators ledding an inflammatory reply. [33].

> The meroterpenes have been isolated from Ganoderma lucidum (lucidumins A-D, lingzhine C, ganocochlearin B, fornicin B, cochlearin I, ganocochlearine A, lucidimine A) by Shuang-Yang and the research team. All meroterpenes were tested for antiinflammatory activity by determining LPS-induced nitric oxide (NO) generation in RAW264.7 macrophages. The outcomes displayed that lucidumins A-D, can significantly inhibit NO production with IC50 values 8.06 \pm 0.51, 10.98 \pm 0.15, 15.49 \pm 0.40, 9.39 \pm 0.21, respectively, which exhibited stronger activity than the positive control L-NMMA [34].

fungus Aspergillus sp. ZL0-1b14. They were assayed for anti- is about the meroterpenes obtained from the plants, one is on the inflammatory macrophages. Aspertetranone D showed an inhibitory effect Five research is focused on the meroterpenes from marine-derived towards IL-6 production with 69% inhibition at 40 µM. [35].

isolated from the species Ulmus davidiana var. Japonica by Choi et. all. Bakuchiol powerfully suppressed the production of nitrogen oxide activated by lipopolysaccharide (LPS), and research group. Plant- related endophytic fungus papers are about prostaglandin E2 (PGE2) in line 264.7 macrophages, without exhibiting cytotoxicity [36]. In another bakuchiol study, it was there is one research on Penicillium sp. from a soil sample and exhibited to inhibit degranulation of human neutrophils in vitro. The same effect was also confirmed in vivo in mice administered with zymosan causing tissue inflammation. It was also determined that bakuchiol fully inhibited myeloperoxidase activity, hence restraining the formation of oxidizing compounds such as hypochlorite. As a result, bakuchiol caused a diminishing in tissue damage caused by hydrolytic enzymes and some oxidizing compounds that existed in human leukocytes [37-38].

Xiao and the research group have found a new meroterpene, psoracorylifol F from Psoralea corylifolia fruits, together with two common meroterpenes (psoracorylifol A and bakuchiol), All the meroterpenes have the capability of an anti-inflammatory agent. three meroterpenes possessed evaluated by TLC bioautography against O2- radicals. They have strong inhibitory activity against Conflicts of Interest: LPS-induced NO production in RAW 264.7 cells with IC50 values varying from 7.71 to 27.63 µM [39].

The COX-1 and COX-2 IC50 for bakuchiol were found to be 14.7 µg/mL and 514 µg/mL, respectively by Chaudhuri & Marchio [40].

Chen et. all. have isolated nine unidentified shikimate-conjugated 2. meroterpenes, as well as nine known compounds from solid cultures of the fungus Guignardia mangiferae. Nine undescribed 3. meroterpenes suppressed nitric oxide (NO) production in LPSactivated RAW 264.7 cells with IC50 values in between 4.7 and 4. 40.0 µM [41].

Zbakh and the research group have studied the anti-inflammatory 5. algal meroterpene 11-hvdroxv-1'-Oactions of the methylamentadione (AMT-E) in a murine model of dextran sodium sulphate (DSS)-promoted colitis. To perform the test, 6. AMT-E was orally subjected daily (1, 10, and 20 mg/kg animal) to DSS-treated mice (3% w/v) for 7 days. AMT-E inhibited body weight loss and colon shortening and efficiently weakened the size of the colonic damage. Likewise, AMT-E increased mucus 7. generation and decreased myeloperoxidase activity (a marker for anti-inflammatory activity). Furthermore, the algal meroterpene reduced the tumor necrosis factor (TNF)- α , interleukin (IL)-1 β , and IL-10 levels, and caused an important reduction of the expression of promotable nitric oxide synthase (iNOS) and 8. cyclooxygenase-2 (COX-2) [42].

Conclusion:

As a result of the literature survey, a limited number of papers are present about the anti-inflammatory research on meroterpenes 10. from different sources. Five of them (Hypericum patulum, Psoralea corvlifolia, Ulmus davidiana var. Japonica, Rhododendron anthopogonoides Maxim., Ganoderma cochlear)

activity in LPS- activated RAW264.7 meroterpenes isolated from mushroom (Ganoderma lucidum). fungus (Aspergillus sp. ZL0-1b14., Pleosporales sp., Penicillium brasilianum., Penicillium chrysogenum J08NF-4. Aspergillus Anti-inflammatory activity has been assayed on bakuchiol terreus GZU-31-1). Marine meroterpenes studies are the investigations belonging to Zbakh and the research group, Makkar & Chakraborty, De Los Reyes, and his co-workers, Zbakh and his Talaromyces amestolkiae YX1, Aspergillus terreus. Additionally, one is the meroterpenes from solid cultures of the fungus Guignardia mangiferae. To measure the anti-inflammatory activity, the production of NO, IL-6, IL-1B, TNF-a in LPS- and prostaglandin E2 (PGE2)-induced RAW 264.7 cells, in LPSinduced THP-1 human macrophages, the inhibitory effect of COX-1 and COX-2, 5- lipoxidase, nitric oxide synthase (iNOS), peroxidase proliferator-activated receptor (PPAR-γ)/NF-κB signaling line and in NF-kB pathway luciferase assay have been performed in the papers we selected. The majority of the meroterpenes in the manuscripts mentioned have extreme antiinflammatory activities in the µM range. It shows us that

The authors declare that they have no conflicts of interest.

References:

- Medzhitov, R. Inflammation 2010: New Adventures of an 1. Old Flame, Cell. 2010,140.
- Schmid-Schonbein, G. W. Analysis of Inflammation, Annu. Rev. Biomed. Eng. 2006, 8, 93–151
- Ross, R. Atherosclerosis—an inflammatory disease, N. Engl. J. Med. 1999. 340, 115–26
- Schmid-Schonbein, G. W., Takase, S, Bergan, J. J. New advances in understanding of the pathophysiology of chronic venous insufficiency. Angiology. 2001. 52 (Suppl. 1), 27-34.
- Engler, R.L., Schmid-Schonbein, G. W., Pavelec, R. S. Leukocyte capillary plugging in myocardial ischemia and reperfusion in the dog. Am. J. Pathol. 1983. 111, 98–111.
- Entman, M.L., Michael, L., Rossen, R.D., Dreyer, W. J., Anderson, D.C., Taylor, A. A., Smith, C. W. Inflammation in the course of early myocardial ischemia. FASEB J. 1991. 5, 2529–37.
- Anselmi, A., Abbate, A., Girola, F., Nasso, G., Biondi-Zoccai, G. G., Possati, G., Gaudino, M. Myocardial ischemia, stunning, inflammation, and apoptosis during cardiac surgery: a review of evidence. Eur. J. Cardiothorac. Surg. 2004. 25, 30411.
- Koistinaho, М., Koistinaho, J. InteractionsbetweenAlzheimer'sdiseaseand cerebral ischemia-focus on inflammation. Brain Res. Brain Res. Rev. 2005. 48, 240-50.
- 9. Iadecola, C., Alexander, M. Cerebral ischemia and inflammation. Curr. Opin. Neurol. 2001. 14, 89-94.
- Jean, W. C., Spellman, S. R., Nussbaum, E. S., Low, W. C. Reperfusion injury after focal cerebral ischemia: the role of inflammation and the therapeutic horizon. Neurosurgery. 1998. 43, 1382-96; discussion 1396-87.

- 11. Del Zoppo, G. J. Microvascular responses to cerebral ischemia/inflammation. Ann. N.Y. Acad. Sci. 1997. 823, 132-47.
- 12. Kontos, C. D., Wei, E. P., Williams, J. I., Kontos, H. A., Povlishock, J. T. Cytochem- ical detection of superoxide in cerebral inflammation and ischemia in vivo. Am. J. Physiol. Heart Circ. Physiol. 1992. 263, H1234-42.
- 13. Del Zoppo G. J., Schmid-Schonbein, G. W., Mori, E., 29. Copeland, B. R., Chang, C-M. Polymorphonuclear leukocytes occlude capillaries following middle cere- bral artery occlusion and reperfusion. Stroke. 1991. 22, 1276–83.
- 14. Suematsu, M., Suzuki, H., Delano, F. A., Schmid-Schonbein, G. W. Theinflam- matory aspect of the microcirculation in hypertension: oxidative stress, leuko- cytes/endothelial interaction, apoptosis. Microcirculation. 2002. 9, 259-76.
- 15. Li, Q., Withoff, S., Verma, I. M. Inflammation-associated 31. cancer: Nf- $\kappa\beta$ is the lynchpin. Trends Immunol. 2005. 26, 318-25.
- 16. Karin, M., Greten, F. R. Nf-κβ: linking inflammation and immunity to cancer development and progression. Nat. Rev. 32. De los Reyes, C., Zbakh, H., Motilva, V., Zubía, E. Immunol. 2005. 5, 749-59.
- 17. Philip, M., Rowley, D. A., Schreiber, H. Inflammation as a tumor promoter in cancer induction. Semin. Cancer Biol. 2004. 14, 433-39.
- 18. Baker, R. G., Hayden, M. S., Ghosh, S. NF-KB, inflammation, and metabolic disease. Cell Metab. 2011. 13, 11 - 22.
- 19. McNelis, J. C., Olefsky, J. M. Macrophages, immunity, and 34. Lu, S-Y., Peng, X-R., Dong, J-R., Yan, H., Kong, Q-H., Shi, metabolic disease. Immunity. 2014 41, 36-48.
- 20. Jia, X., Wu, Y., Lei, C., Yu, Y., Li, J., Li, J., Hou, A. Hyperinoids A and B, two polycyclic meroterpenoids from Hypericum patulum. Chinese Chemical Letters. 2020 31, 1263-1266.
- 21. Qin, Fu-Y., Yan, Y-M., Tu, Z-C., Cheng, Y-X. (±) Gancochlearols A and B: cytotoxic and COX-2 inhibitory meroterpenoids from Ganoderma cochlear. Natural Product Research. 2020. 34, 16, 2269-2275.
- 22. Shi, Q., Li, T-T., Wu, Y-M., Sun, X-Y., Lei, C., Li, J-Y., Hou, A-J. Meroterpenoids with diverse structures and anti- 36. Choi, S-Y., Lee, S., Choi, W., Lee, Y., Yo, J., Ha, T-Y. Rhododendron inflammatory activities from anthopogonoides. Phytochemistry. 2020. 180, 112524.
- 23. Tang, Y., Chen, X., Zhou, Y., Zhao, Min., He, J., Liu, Y., Chen, G., Zhao, Z., Cui, H. Furanaspermeroterpenes A and 37. Ferrandiz, M. L., Gil, B., Sanz, M. J., Ubeda, A., Erazo, S., B, two unusual meroterpenoids with a unique 6/6/6/5/5 pentacyclic skeleton from the Marine-derived fungus Aspergillus terreus GZU-31-1. Bioorganic Chemistry. 2021. 114, 10511.
- 24. Zbakh, H., Zubía, E., De los Reyes, C., Calderón-Montaño, 38. Jafernik, K., Halina, E., Ercisli, S., Szopa, A. Characteristics J. M., López-Lázaro, M., Motilva, V. Meroterpenoids from the Brown Alga Cystoseira usneoides as Potential Anti-Inflammatory and Lung Anticancer Agents Mar. Drugs. 2020.18, 207.
- 25. Liu, Z., Liu, H., Chen, Y., She, Z. A new anti-inflammatory meroterpenoid from the fungus Aspergillus terreus H010. Natural Product Research. 2018. 32, 22, 2652-2656.
- 26. Liu, S., Su, M., Song, S. J., Hong, J. Chung, H. Y., Jung, J. H. An anti-inflammatory PPAR-gamma agonist from the jellyfish derived fungus Penicillium chrysogenum J08NF-4. J. Nat. Prod. 2018. 81, 356-363.
- 27. Zhang, J., Yuan, B., Liu, D., Gao, S., Proksch, P., Lin,W. BrasilianoidsA-F, new meroter penoids from the sponge

associated fungus Penicillium brasilianum. Front. Chem. 2018. 6, 314–326.

- 28. Chen, C. J., Zhou, Y.Q., Liu, X. X., Zhang, W. J., Hu, S. S., Lin, L. P., Huo, G. M., Jiao, R. H., Tan, R. X., Ge, H. M. Antimicrobial and anti-inflammatory compounds from a marine fungus Pleosporales sp. Tetrahedron Lett. 2015. 56, 6183-6189.
- Jiang, M., Wu, Z., Guo, H., Liu, L., Chen, S., A Review of Terpenes from Marine-Derived Fungi: 2015–2019 Mar. Drugs. 2020. 18, 321.
- 30. Etoh, T., Kim, Y. P., Tanaka, H., Hayashi, M. Antiinflammatory effect of berkeleyacetal C through the inhibition of interleukin-1 receptor-associated kinase-4 activity. European Journal of Pharmacology. 2013. 698, 435-443.
- Chen, S., Ding, M., Liu, W., Huang, X., Liu, Z., Lu, Y., Liu, H., She, Z. Anti-inflammatory meroterpenoids from the mangrove endophytic fungus Talaromyces amestolkiae YX1. Phytochemistry. 2018. 146, 8e15.
- Antioxidant and Anti-inflammatory Meroterpenoids from the Brown Alga Cystoseira usneoides. J. Nat. Prod. 2013. 76, 621-629.
- 33. Makkar, F., Chakraborty, K. Antioxidant and antiinflammatory oxygenated meroterpenoids from the thalli of red seaweed Kappaphycus alvarezii. Medicinal Chemistry Research. 2018. 27, 2016–2026.
- Q-Q., Li, D-S., Zhou, L., Li, Z-R., Qiu, M-H. Aromatic Ganoderma lucidum constituents from and their neuroprotective and anti-inflammatory activities. Fitoterapia. 2019.134,58-64.
- 35. Wang, Y., Qi, S., Zhan, Y., Zhang, N., Wu, A-A., Gui, F., Guo, K., Yang, Y., Cao, S., Hu, Z., Zheng, Z., Song, S., Xu, Q., Shen, Y., Deng, X. Aspertetranones A-D, Putative Meroterpenoids from the Marine Algal-Associated Fungus Aspergillus sp. ZL0-1b14. J. Nat. Prod. 2015. 78, 2405-2410.
- Isolation and anti-inflammatory activity of bakuchiol from Ulmus davidiana var. japonica. J Med Food. 2010. 13, 4, 1019-1023.
- Gonzalez, E., Negrete, R., Pacheco, S., Payaa, M., Alcaraz, M. J. Effect of bakuchiol on leukocyte functions and some inflammatory responses in mice. J Pharm Pharmacol. 1996. 48, 9, 975–980.
- of bakuchiol- the compound with high biological activity and the main source of its acquisition - Cullen corylifolium (L.) Medik. Natural Product Research. 2021. 35, 24, 5828–5842.
- 39. Xiao, G., Li, X., Wu, T., Cheng, Z., Tang, Q., Zhang, T. Isolation of a new meroterpene and inhibitors of nitric oxide production from Psoralea corylifolia fruits guided by TLC bioautography. Fitoterapia. 2012. 83, 1153-1557.
- Chaudhuri, R. K., Marchio, F. Bakuchiol in the Management 40. of Acne-affected Skin, Cosmetics & Toiletries® magazine. 2011. 126, 7.
- 41. Chen, K., Chen, C., Liu, X., Sun, We., Deng, Y., Liu, J., Wang, J., Luo, Z., Zhu, H., Zhang, Y. Terpene- Shikimate conjugated meroterpenoids from the endophytic fungus

J Pharmacy and Drug Innovations



Guignardia mangiferae. Phytochemistry. 2021. 190, 112860.

42. Zbakh, H., Talero, E., Avila, J., Alcaide, A., De los Reyes, C., Zubía, E., Motilva, V. The Algal Meroterpene 11-Hydroxy-1'-O-Methylamentadione Ameloriates Dextran Sulfate Sodium-Induced Colitis in Mice. Mar. Drugs. 2016. 14, 149.